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Title: In vivo stability of WS1321 AASD strains carrying HIV peptide

Dates:

Purpose: Determine the number of bacteria recovered from the spleens of orally infected BALB/c mice. Compare the levels of each <sup>bacterial</sup> strain to that of the parent strain.

Strains: Constructed by Art Brantstrom are:

WS1321AASD, PAB102

WS1321AASD, PAB103

WS1321AASD, PAB102::gag

WS1321AASD, PAB103::gag

WS1321AASD, PAB102::rif

WS1321AASD, PAB103::rif

Also

WS1321 - plasmid - Sal. typhimurium strain

See to explain purpose  
of construction of these 2  
strains.

Note: A previous experiment conducted indicated by Day 6 after <sup>oral</sup> infection fewer numbers of strain WS1321, PAB103 were recovered from the spleens. ← Lab Notebook #3 (Mouse work)

Experiment: (Full procedure documented in) Lab Notebook #3 (Mouse work)

- strains were grown to an OD. of  $\approx$  1.8 - 1.0
- strains to concentrate into DPBS (Some cultures resuspended in 0.35 ml. strains into 0.5 ml.)
- cultures were diluted & plated to determine <sup>approx</sup> bacterial fed.
- 40ul fed to mice using 200ul pipet.
- mice are to be sacrificed on Days 4, 7 & 11 post-feeding.  
3 mice each strain each timept.

amt. given to mice: Between  $3 - 4 \times 10^9$  in 40ul.

Exact #'s are in Book #3

\* Spleens homogenized in 1ml HBSS 0.1ml original placed in duplicate

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PROJECT Salmonella Vaginale Canis

Results: ave/spore = ave of  $3 \times 1/0.5 \times 2$  ml dil.

Strain	Day	ave/spore	Day	ave/spore	Day	ave/spore			
							total	total	total
WS1321 #1	4	90,101	7	125,122	11	95,831			
# 2	131,46	> 1697	98,71	> 2097	11	111,103	>	1690	
# 3	70,71		107,97			65,51			
WS1321 #1	44,37		8,9			63,100			
PAB102 #2	14,86	> <del>265</del>	92,135	> 1317	19,17	> 1123			
# 3	10,66	1160	73,78			74,64			
WS1321 PAB103 #1	5,4		35,46	> 70		97,57			
# 2	2,9	> 10	59,45	> 77	2,3	> 1283			
# 3	0,0		26,21			104,122			
WS1321 PAB103 #1	0,0		0,0			3,4			
# 2	0,0	> 0	18,19	> 507	0,0	> 46			
# 3	0,0		54,61		51				
WS1321 PAB103 #1	0,0		0,0			0,0			
# 2	0,0	> 0	0,0	> 0	3,0	> 10			
# 3	0,0		0,0		0,0				
WS1321 PAB103 #1	0,0		0,0			1,2			
# 2	0,0	> 0	0,1	> 27	6,9	> 63			
# 3	0,0		0,0		6,0				
WS1321 PAB103 #1	0,0		0,0			1,0			
# 2	0,0	> 0	0,0	> 0	4,4	> 53			
# 3	0,0		0,0		3,4				
> 20 not detectable									

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Cloning and Testing HIV genes under the control of the pgC promoter.

Art Blanckom constructed plasmid pAB104, which is plasmid pAB103 w/ the pgC promoter replacing the lac promoter.

\* All data in Notebook #1 Digestions & Ligations

Using directed cloning techniques a 227 Kb fragment encoding Vif from HIV was cloned into pAB104

Britten:

pAB104	5ul
B buffer	0ul
BamH1	1ul
dH2O	1ul
	20ul

pAB104: Vif	10ul
B buffer	2ul
BamH1	1ul
dH2O	7ul
	20ul

↓ 1 hr. 37°C

- add to each & continue at 37°C  
for several hours.

Above	20ul
H buffer	3.2ul
Sal I	2ul
dH2O	14.8ul
	40ul

- + Each mix was run on a 1.2% low melt agarose gel. 1/2 mix on back side of the gel. With the gel was stained w/ EtBr after electrophoresis. The appropriate bands were sliced from the bottomed side.
- Each band was Prep-A-Gene purified from through a Millipore column (a minicolumn) + resuspended in 25ul TE for ligation.

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PROJECT Salmonella Vacuole Carriers

NOTE NO. \_\_\_\_\_

continued From Page 3Digestion:

Control + pAB104 cut, purified & digested  
 10X buffer 5 μl  
 10mM ATP 4 μl  
 pAB104 cut, purified 12.5 μl  
 T4 liga~~se~~se 2 μl  
 dH<sub>2</sub>O 26.5 μl  
 50 μl

pAB104 + vif  
 10X buffer 5 μl  
 10mM ATP 4 μl  
 pAB104 12.5 μl  
 Vif 2.5 μl  
 T4 Liga~~se~~se 2 μl  
 dH<sub>2</sub>O 15 μl  
 50 μl

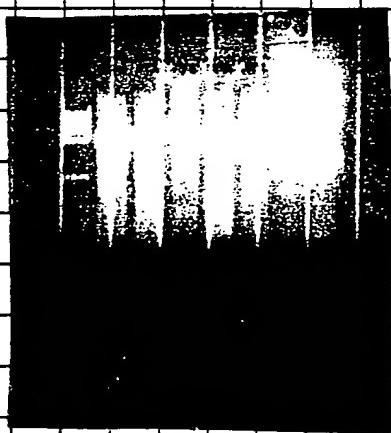
↓  
 15°C, overnight in PCR machine  
 15 min 12 min gel

- Ethanol prec. & electroporate into XG097ΔASD

	Time (min)	colonies	Ligation Mix
pAB104	13.7	0	[Redacted]
pAB104::vif	14.8	many.	[Redacted]

5 positives were selected for digestion to look for inserts:

- Wilmot Miniprep - purify DNA
- Cut w/ BamH - SstI
- Run a gel.



all five appear to have inserts.

All five were prepared for Western Blot examination of expression levels. #10 were electroporated into JB501Tn10::ASD for placement into 1B21ASD.

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PROJECT

Salmonella Vaccine Carriers

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Title: Macrophage Assay of pAgC construct

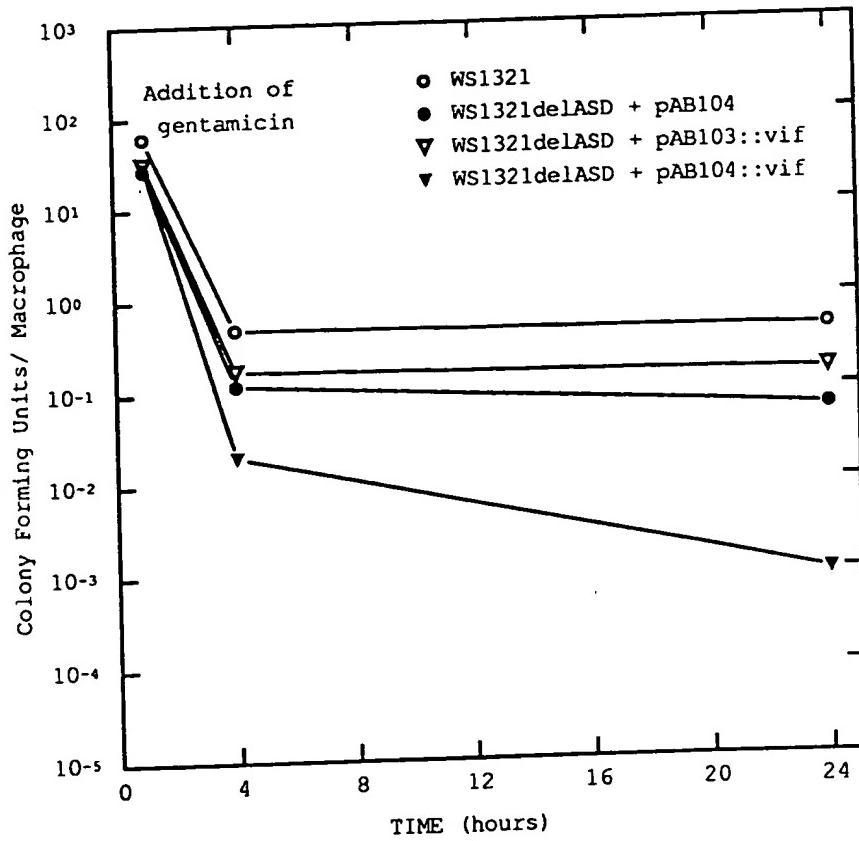
Purpose: Determine whether pAgC promotes increases expression in murine Macrophages

Notebook pag C #1 full experimental protocol.

Assay based on protocol of O'Brien et al.

CFU/Macrophage & Expression via Western blot were assayed.

Results:



Western in pag C notebook - Shows expression  
from pAgC (pAB104::vif only).

Slides of Macrophage infected in Petri dish - notebook - pag C #1  
Experiment Repeated

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Title: Growth of WS1321 Δ ASP + pAB104 in Minimal Media  
 Supplemented w/ 0.1% Glucose

Notebook pag C #1

Repeated

Protocol:

- Single colony of each strain into 5ml of LB or MEM
- Grow 3-4.5 hrs.
- Concentrate & resuspend in dissociation buffer, vortex  
freeze → Thaw
- Determine protein concentrations
- Run Western to compare LB & MEM expression  
of lacZ & pagC promoters.

Results: (Notebook pag C #1)

— 3/16 — EVP.

Bacterial Counts also determined

Lane # 10. Marker

&lt; 9. WS1321

(LB) 8 pAB103::lacZ

&lt; 7 pAB104

- 6 pAB104::lacZ

5 WS1321

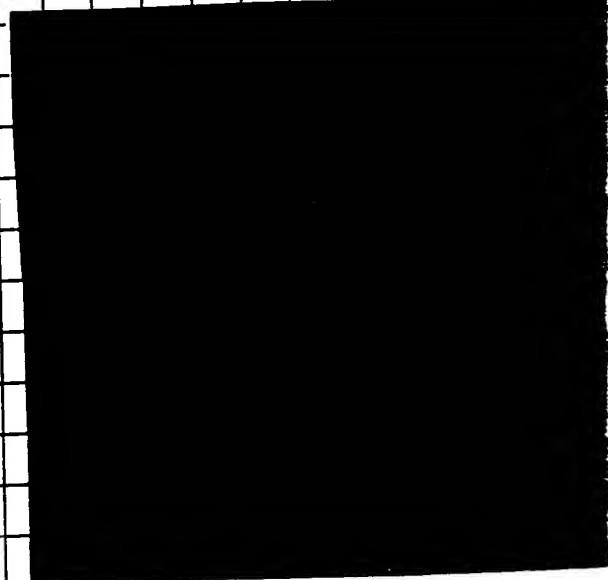
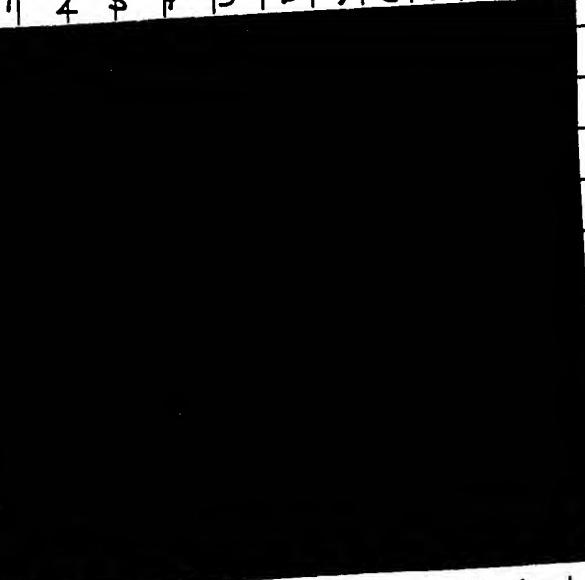
4 pAB103::lacZ MEM

3 pAB104

2 pAB104::lacZ

1 2 3 4 5 6 7 8 9 10

1 2 3 4 5 6 7 8 9 10



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Title: Mouse Experiment page Controls  
 Strains Tested: WS1321  
 WS1321 A ASD+ pAB103::vif  
 WS1321 A ASD+ pAB104  
 WS1321 A ASD+ pAB104::vif

Purpose: Determine the number of bacterial colonies surviving mouse passage & expression of vif gene after passage.  
 Mice sacrificed on days 3, 7 & 10

Amount of Bacteria Fed to Mice

40ul

WS1321	$4.4 \times 10^9$
pAB103::vif	$4.2 \times 10^9$
pAB104	$6.9 \times 10^9$
pAB104::vif	$6.4 \times 10^9$

Results: Experimental problem: Apparently mice were mixed by animal handlers.  $\leftarrow$  (Based on) Western blot analysis

Donata L. Segmire

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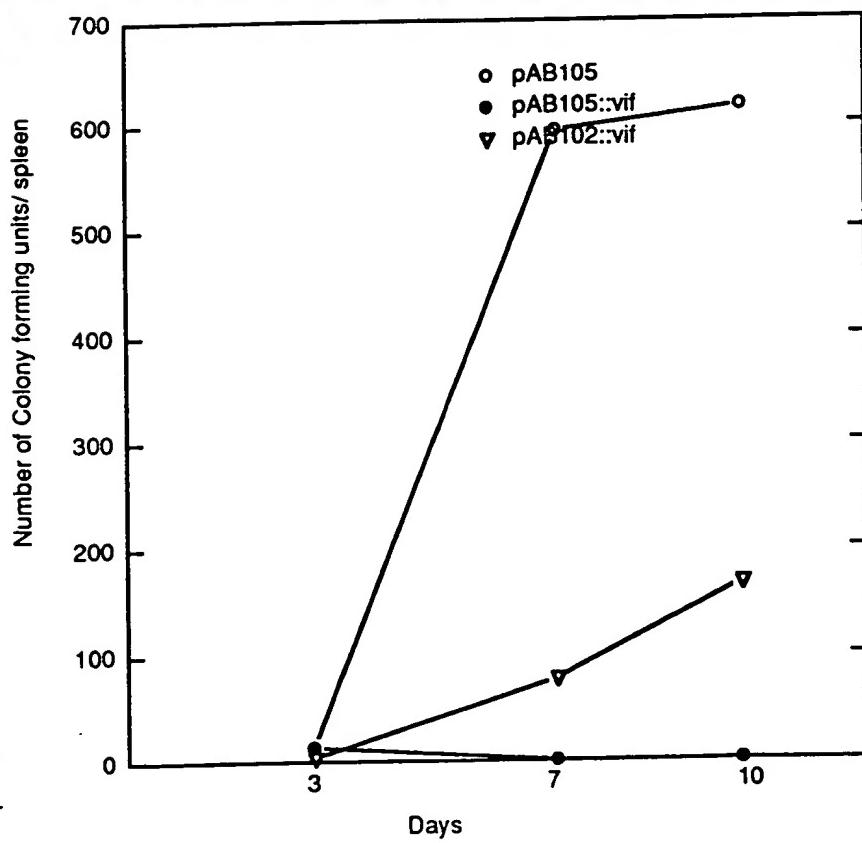
Title: Construction of WS1321AASD strains carrying pAB105 & pAB105::vif for mice feeding.

Purpose: pAB105 & pAB105::vif plasmids were constructed by recombination. In this plasmid the asd gene is from *E. coli*. We believe strains carrying plasmids to this asd gene are healthier.

5/12 - Mice fed the following diet in amount  
40ul

WS1321AA5D + pAB105  $3.6 \times 10^9$   
WS1321AASD + pAB105::vif  $2.72 \times 10^9$   
WS1321AASD + pAB102::vif  $6.4 \times 10^9$

Results: CFU/spleen + expression of vif from recovered colonies



Results of Western  
Only pAB102::vif  
showed significant  
expression.

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Title: Construction of pAB105::ENV113

Purpose: Of the HIV genes we worked with only ENV has mapped cDNA sequences of a BALB/c mice.

After several attempts: → it was determined that the entire ENV gene isn't stable in our pGK plasmid. Various deletions resulted after each cloning attempt. Ended July

Will try smaller fragment which has been constructed by Art Ransdorn

End

~~Denote R. Segmore~~

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Title : Summary of Human Monocyte & S.typhi  
Interaction Experiments. July

Note : This work has been in progress for some time.  
This entry is a summary of the work completed  
& a list of work needed to be done for a problem.

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Initial CTL Assay

Purpose: To standardize non-radioactive CTL assay from Biomega. Testing various fetal calf serum lots.

Testing:

#1 FBS certified (Cow LDH ≈ 470 units) Gibco  
Lot # 40K0242

#2 Nyclone

5ml of #1 or #2

45ml of RPMI w/out Phenol red.

50 ml

- add 50ul to each well
- add 50ul of Control + LDH to 6 wells
- add 10ul of Cytosol solution
- add 50ul of Substrate Mix (12ml of Assay Buffer to bottle of Substrate Mix.) Cover plate w/ foil
- \* incubate at RT for 30'
- Add 50ul of Stop Solution
- Remove large bubbles. Recal Ab at 490.

Note - better control RPMI 1640 w/out serum.

Results: #1 best #2 cannot be used - completely red.  
Aug 16 1994

## Control Testing Maximum &amp; Spontaneous Release of PS15

#1 Group - RPMI

#2 Group - Media #2 - CTL Assay Media

Note: Cells are healthy. This is a must.

$5 \times 10^3$  &  $1 \times 10^4$  cells were tested for spontaneous (maximun) release.

RPMI w/out phenol red & Media #2 w/out phenol red. were used as control backgrounds.

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Results:

RDM 1  
<sup>start Min.</sup>  
<sup>start Min.</sup>  
 $5 \times 10^3 \quad 0.479 \quad 0.158 \quad \times 3.5$   
 $1 \times 10^4 \quad 0.748 \quad 0.158 \quad \times 4.7$

Media + 2  
<sup>start Min.</sup>  
 $0.537 \quad 0.153 \quad \times 3.5$   
 $1.083 \quad 0.150 \quad \times 7.22$

$$\text{RDI} = 0.074$$

$$\text{Media} = 0.162$$

$\downarrow 3\% \text{ FBS}$

$$\text{Gibco } 10\% = 0.326$$

Assay looks like it should work.

Final runs for CTL Proliferation & Cytokine  
 Assay in Shigella 2a ENVTS  $\rightarrow$  experiment.

Comments uncalculated by Coley Mallett.  
 ca

Above data contained in Shigella Cpp/Hela/HepG2  
 Notebook.

Based on finds by Cut Brantum a pUC18 plasmid  
 encoding ENVTS-A, present is stably maintained  
 in Shigella flexneri 2a without kanamycin.

Cut Brantum has checked all strains used  
 for this study for Congo Red binding; I believe  
 + plasmid content.

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(Cloning of ENVI<sub>B</sub>-A<sub>1</sub> plc ~...+ into pAB105)

Purpose: This ENVI<sub>B</sub>-A<sub>1</sub> fragment contains the relevant V3 loop, CTL epitope + major Ab epitope. Furthermore, it appears to be more stable in Puc8.

Outline of cloning procedure in rough notes: Pg C Notebook

Results: Appears to be cloned. Several weeks of problems w/ ligase!

Several mAb used for detection of expression

1. human sera (Ab's)
2. human sera (TD)
3. mAb R 9-2
4. V3 (HTB-V3-13) — only one that is good!

Mabs Run

1. Oug 1 well protein  
Comparing pAB102::ENVI<sub>B</sub>-A<sub>1</sub> to pAB105::ENVI<sub>B</sub>-A<sub>1</sub>  
(plac) (pAgc)

Repeated Results:

- plac lysates from Art for 11-28-94  
not enough protein loaded

- pAgc expression is clearly higher than plac.

Need to repeat w/ fresh Ab, but all looks good.  
Set up for mouse testing.

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Stargate CMI Response

59 mice total: 45<sup>th</sup> floor

Purpose: Test for CTL activity to EN1 epitope.

Mice sacrificed Sept. 6: Note: animals look ruffled.  
- 2 S. flexneii 2a died.

Cells prepared for CTL, proliferation &amp; cytokine assays.

Concept dropped

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## PROJECT Shigella as a DNA Carrier

Previous background experiments appear in Notebooks I & II.

As of , to extend this concept we will step antibiotic treated BHK cells which have been infected w SC6002(pCMVp). The idea is that antibiotic treatment will kill the intracellular bacteria. The dead bacteria will release the pCMVp plasmid into the cytoplasm for transcription either done by the BHK cell

This entry is based on a conversation with Tony Daboff on [redacted]. This entry was made on [redacted] by [redacted].

- This entry was  
~~Doreck L. Simeone~~

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**Rr and Understood By**

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Date

Esther Hartman

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